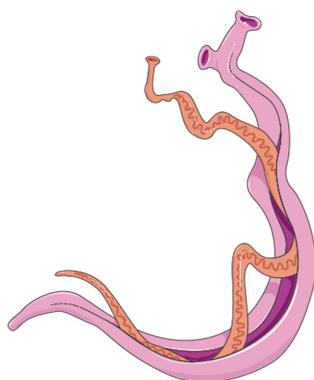


Schweizerische Fachgesellschaft für Tropen- und Reisemedizin FMH
Société Suisse de Médecine Tropicale et de Médecine des Voyages FMH
Società Svizzera di Medecina Tropicale e di Viaggio FMH
Swiss Society of Tropical and Travel Medicine FMH



Swiss Tropical and Public Health Institute
Department of Medicine
Medical and Diagnostic Services
National Reference Centre for Imported Parasitic Diseases

Schistosomiasis Treatment Recommendations 2020



Schistosomiasis	Parasite	1st-line	Comments
(acute phase)	characteristics	Diagnostics	treatment
<i>S. haematobium</i>	Definitive host: <i>S. man.</i> : humans, primates, rodents; <i>S. h.</i> : humans, primates, pigs, buffalos;	• Eosinophilia ^x • PCR (blood) • (Serology ^x)	• Acute schistosomiasis (AS; 'Katayama fever') is an immune-mediated serum sickness-like hypersensitivity reaction to migrating and maturing schistosomulae. AS is usually only seen in immunologically naïve patients (travellers) and not in individuals from endemic regions.
<i>S. mansoni</i>	<i>S. i., S. g.</i> : humans only;	• (Circulating antigen in blood/urine [CCA, CAA] ^x)	• ^x The absence of eosinophilia, circulating parasite antigen or a negative serology does not rule out AS! Eosinophilia and seroconversion may be delayed for up to 3 weeks following the onset of symptoms. Circulating antigens (CCA/CAA) become detectable in blood/urine ≥ 3 weeks post-infection. PCR is an emerging diagnostic tool in AS (see comment ^ø p. 107).
<i>S. intercalatum</i>	<i>S. j.: various domestic and wild mammals, humans;</i>	Δ Praziquantel 60 mg/kg/d p.o. (in 2 or 3 doses) on day 3 after initiation of treatment ^ø	• ^ø The use of PZQ in AS remains controversial. PZQ is ineffective on young (7–28 days old) schistosomulae and thus, its efficacy in AS is limited. In addition, PZQ treatment is associated with paradoxical (Jarisch-Herxheimer-like) reactions in 40–59% of AS cases [2,3] (see comment* below). Some experts advocate giving PZQ in AS (mostly because of the fear of acute neuroschistosomiasis; see below), others advocate delaying its use until the chronic phase of infection has been reached [4]. The concomitant use of steroids may prevent/attenuate PZQ treatment-related paradoxical reactions.
<i>S. guineensis</i>	<i>S. mekongi</i>	\circ Ivermectin ^ø 200 µg/kg p.o. single dose	• ^ø Consider ruling out concomitant neurocysticercosis (by serology) in patients from endemic regions before giving PZQ, as this may provoke seizures [1].
<i>S. japonicum</i>	<i>S. malayensis</i>	+ add a complete course of Praziquantel (see next page)	• ^ø Artemisinins show some activity against young schistosomulae. Adding an artemisinin to praziquantel may be beneficial.
<i>S. mekongi</i>	<i>S. mattheei</i>	Follow-up: see below	• ^ø When giving high dose steroids, consider adding ivermectin to cover for possible co-infection with strongyloidiasis if patients are from high-endemic regions or strongyloides screening serology is positive.
<i>S. malayensis</i>	Clinical picture: Fever, dry cough, fatigue, headache, myalgia, arthralgia, abdominal pain, diarrhoea, rarely: periorbital oedema, urticaria/rash, encephalitis, myocarditis	Route of infection: fresh water contact, penetration of skin by cercariae (500 µm), development into 'schistosomulae' and then into adult worms (10–20 mm)	• ^ø In some patients relapsing forms of AS have been observed (~15 days after the primary symptomatic episode).
<i>S. mattheei</i>		Incubation period: 14–84 days	
		Prepatent period: 4–6 weeks (<i>S. m.</i>) ^y – 3 months (<i>S. h.</i>) ^x	

^x 3 months is considered to be the maximum prepatent period in most cases. However, in rare cases the acute phase might be much longer (>5 months) and severe treatment-induced paradoxical reactions may occur very late after infection. A high eosinophil count in the absence of detectable eggs should (irrespective of elapsed time since exposure) always raise the suspicion of an acute phase infection and the risk of treatment-induced paradoxical reactions [5].

[1] Torres JR. Use of praziquantel in populations at risk of neurocysticercosis. Rev Inst Med Trop São Paulo 1989;31:290.

[2] Grandière-Pérez L et al. Efficacy of praziquantel during the incubation and invasive phase of *S. haematobium* schistosomiasis in 18 travelers. Am J Trop Med Hyg 2006;74:814–8.

[3] Bottieau E et al. Imported Katayama fever: clinical and biological features at presentation and during treatment. J Infect 2006;52:339–45.

[4] Jauréguiberry S et al. Acute schistosomiasis, a diagnostic and therapeutic challenge. Clin Microbiol Infect 2010;16:225–31.

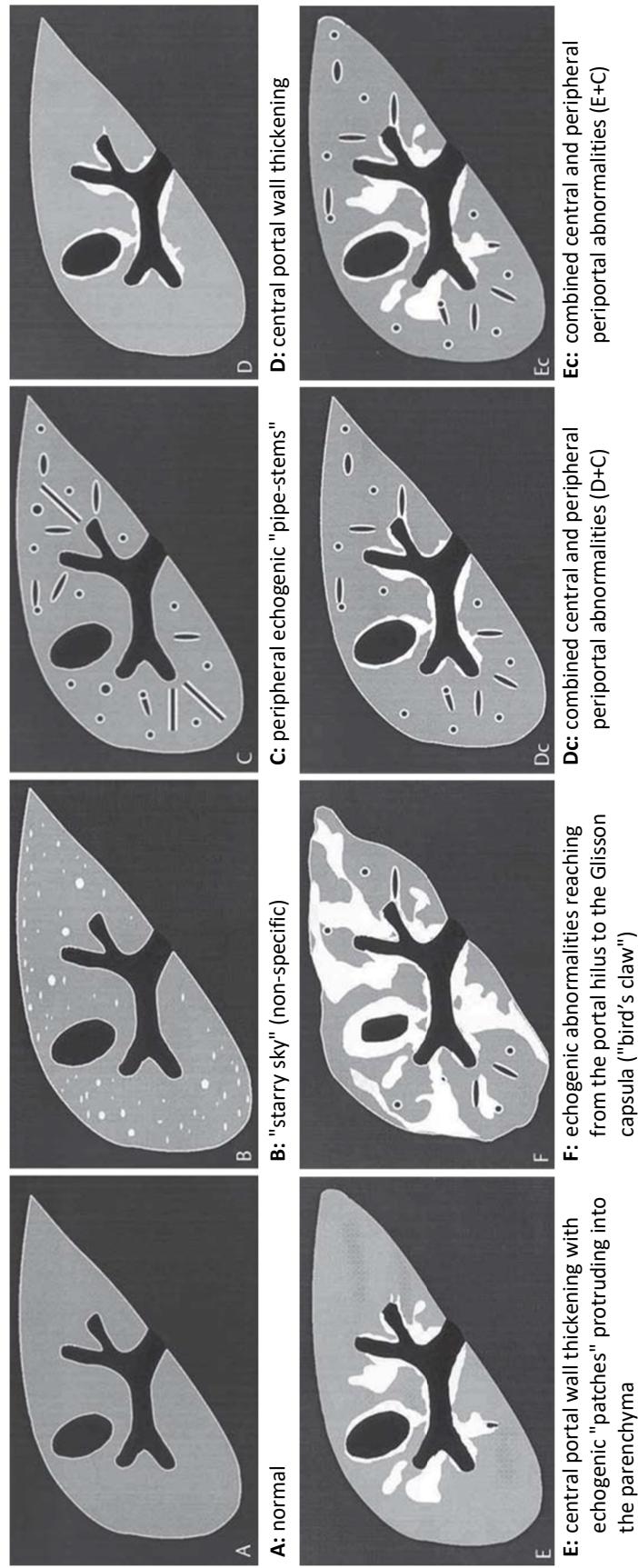
[5] Neumayr A et al. Acute febrile respiratory reaction after praziquantel treatment during asymptomatic late form of acute schistosomiasis. J Travel Med 2012;19:264–7.

Schistosomiasis	Parasite (chronic phase)	Characteristics	Diagnostics	1st-line treatment	2nd-line treatment	Comments
<i>S. haematobium</i> (genitourinary S.)	see above	Egg detection in urine (<i>S. h.</i>) or faeces (other <i>S.</i> spp.)	Praziquantel*	In the case of <i>S. h.</i>, <i>S. man.</i>, <i>S. i.</i>: treatment failure re-treat with Praziquantel	<ul style="list-style-type: none"> Consider ruling out concomitant neurocysticercosis (by serology) in patients from endemic regions before giving PZQ, as this may provoke seizures [3]. Cure rates of a 40–60 mg/kg single dose range from 60–90% [4,5] (see comment on the dosage of PZQ on next page). 	
<i>S. mansoni</i>	life expectancy of adult parasite: 3–5 years	• Serology	on day 0 and on day 21–30 [†]	Oxamniquine	<ul style="list-style-type: none"> PZQ is ineffective on young (<30 days old) schistosomulae → treatment needs to be repeated after all worms have reached maturity. If no (re)exposure to potential source of infection in the past 3 months (max. prepatent period), no need for a time lag between the 2 doses. 	
<i>S. intercalatum</i>	(Note: although in the past the longevity of the parasite, of the parasite, ranging from 18 to 37 years, has been stressed, the average worm lifespan is 3–5 years)	• PCR (urine/faeces)	25 mg/kg p.o. TID on day 0 and on day 21–30 [§]	Oxamniquine: only effective against <i>S. mansoni</i> ; efficacy of a single dose >80%, resistance reported in Brazil and Kenya; limited availability.		
<i>S. guineensis</i>		• Circulating antigen [CCA, CAA] in blood/urine*	75 mg/kg/d p.o. (in 2 or 3 doses) on day 0 only: Metrifonate;	Oxamniquine: only effective against <i>S. mansoni</i> ; efficacy of a single dose >80%, resistance reported in Brazil and Kenya; limited availability.		
<i>S. japonicum</i>		• Abdominal sonography	and on day 21–30 [†]	• Unlike travellers, patients from endemic regions often have a high parasite burden predisposing them to developing sequelae (portal hypertension, bladder cancer) → screen respectively.		
<i>S. mekongi</i>		(classification of hepatointestinal <i>S.</i>)	high toxicity).	<ul style="list-style-type: none"> * Female genital schistosomiasis may affect the entire genital tract manifesting as pain, contact bleeding, and infertility. S.h. may occasionally cause hepatic complications as well. 		
<i>S. malayensis</i>		(classification of hepato schistosomiasis p. 105)		<ul style="list-style-type: none"> In the absence of concomitant hepatic morbidity, impairment of hepatic function/hepatocellular failure is rare even in long-standing infections as schistosomiasis is characterized by liver fibrosis rather than cirrhosis. Antigen detection is indicative of active infection/treatment failure and would thus be more useful than serology. However, sensitivity is still unclear (especially in travellers in whom parasite density is mostly low) and currently only one point-of-care (POC)/rapid diagnostic test for CCA detection in urine is commercially available. For details, see comment 'laboratory testing / screening for schistosomiasis...' p. 107. 		
<i>S. mattheei</i> (hepatointestinal S.)		• (Egg detection in mucosal biopsy)		<ul style="list-style-type: none"> ▲ Transjugular portosystemic shunt (TIPS) has been successfully used for prophylaxis of variceal bleeding [6,7]. 		
Clinical picture: Asymptomatic ↔ genitourinary S.: haematuria, dysuria, obstructive uropathy, bladder malignancy, female infertility*		Note: <i>A desensitization protocol for patients allergic to, but requiring PZQ treatment, has been published [1]</i>				
		Recommended testing strategy: p. 107 If eggs are detected in faeces, check urine and vice versa to exclude double infection				
		Follow-up: see p. 108				

Comment on the praziquantel dosage recommendations in *S. haematobium* and *S. mansoni* infection:

The recommended regimens vary widely, ranging from 40 mg/kg single dose to 40 mg/kg for 3 consecutive days. The WHO recommended single dose 40 mg/kg regimen is derived from studies conducted in endemic regions, where cyclic mass drug administration primarily aims at parasite reduction rather than parasite eradication. In addition, the treatment outcome in these studies was assessed by investigating a single urine- or stool sample, which has low sensitivity. Considering that (i) the failure rate in *S. mansoni* treatment studies decreases with increasing doses (20mg/kg: 51%; 30mg/kg: 35%; 40mg/kg: 23%; 60mg/kg: 17%)[1], (ii) that PZQ is generally very well-tolerated, and (iii) that in individual treatment parasite eradication rather than parasite reduction is the goal, several specialists recommend to use higher dosages than recommended by WHO (e.g. the German Society for Tropical Medicine recommends treating with 40 mg/kg for 3 consecutive days [2]).

[1] Danso-Appiah A et al. Drugs for treating *S. mansoni* infection. Cochrane Database Sys Rev 2008;3:CD000528. [2] <https://www.dtg.org/empfehlungen-und-leitlinien.html>

Ultrasound classification of hepatic (periportal) fibrosis associated with chronic hepatointestinal schistosomiasis:

Good review on sonographical aspects of schistosomiasis: Richter J et al. Ultrasound assessment of schistosomiasis. Z Gastroenterol 2016;54:653-60.

Comment on MRI assessment of schistosomiasis-related hepatic fibrosis: It has been tried to assess hepatic (periportal) fibrosis by MRI and to correlate MR and ultrasound imaging patterns. However, the correlation between MRI and ultrasound patterns is poor (Silva LC et al. *Schistosoma mansoni*: magnetic resonance analysis of liver fibrosis according to WHO patterns for ultrasound assessment of schistosomiasis-related morbidity. Mem Inst Oswaldo Cruz. 2010;105:467-70).

Treatment of schistosomiasis in pregnancy and during breast feeding:

To date, no praziquantel-related mutagenic, teratogenic or embryotoxic effects have been observed. Although human data are limited, a strong argument for treatment is that active infection poses a threat to the mother as well as to the foetus. For details and references on the use of praziquantel in pregnancy and during breast feeding; see drug profile p. 164.

Neuroschistosomiasis		Diagnosis	Treatment	Comments
Schistosomal myeloradiculopathy	see above +	+ Methylprednisolone 15 mg/kg/day (max. 1 g) for 5 days followed by Prednisolone 1 mg/kg/day for up to 6 months	Adjunct supportive treatment: - Pain therapy - Physiotherapy - Urinary bladder catheterization - Early diagnosis and treatment of urinary tract infections - Decubitus prophylaxis - Psycho-/occupational therapy	<ul style="list-style-type: none"> • Neuroschistosomiasis (NS) is rare, but may be seen in up to 2–5% of infected patients. • Neuroschistosomiasis results from ectopic egg deposition in the CNS and neurological symptoms result from the host's inflammatory response to the antigens released by parasite eggs. • Eggs may reach the CNS by embolisation from the portal, mesenteric, and pelvic venous system or as a result of the anomalous migration of adult worms, followed by <i>in situ</i> egg deposition. The occasional finding of adult <i>S. mansoni</i> within leptomeningeal veins and the presence of eggs grouped together in a confined area also suggest migration of the parasite to the CNS.^{x1} • In clinical practice, a presumptive diagnosis of NS is made based on clinical, laboratory, and epidemiological data (and once other differential diagnoses have been ruled out), as confirmation by biopsy is mostly not feasible. • CNS involvement in schistosomiasis can occur in acute primary infection.
All S. spp. may infect the CNS; spinal cord involvement is most frequently caused by <i>S. haematobium</i> and <i>S. mansoni</i>	<ul style="list-style-type: none"> • CSF: - Eosinophilia (40% of cases) - Raised protein level (90% of cases) - Raised mononuclear cell count (Biopsy)^{y1} 	<ul style="list-style-type: none"> + Praziquantel 60 mg/kg/day p.o. in 2–3 doses + Ivermectin 200 µg/kg p.o. single dose^o 	<p>If signs of intracranial hypertension are present:</p> <p>In the absence of signs of intracranial hypertension:</p> <p>Prednisolone 1 mg/kg/day for 2 months (+ tapering)</p> <p>+ Praziquantel 60 mg/kg/day p.o. in 2–3 doses</p> <p>on days 30, 60 and 90</p> <p>+ consider empirically adding Ivermectin 200 µg/kg p.o. single dose^o</p> <p>+ anticonvulsive treatment if patient presents with seizures</p>	<p>If signs of intracranial hypertension are present:</p> <p>Primarily neurosurgical approach and removal of lesion(s) followed by medical treatment</p>
All S. spp. may infect the CNS; cerebral schistosomiasis is most frequently caused by <i>S. japonicum</i>				<p>Good reviews on Neuroschistosomiasis:</p> <ul style="list-style-type: none"> [1] Vale TC et al. Neuroschistosomiasis mansoni: literature review and guidelines. <i>Neurologist</i> 2012;18:333–42. [2] Carod Artal FJ. Cerebral and spinal schistosomiasis. <i>Curr Neurol Neurosci Rep</i> 2012;12:666–74.

Laboratory testing / screening for schistosomiasis in travellers and migrants:

I. Screening of asymptomatic persons (Note: screening should not be done earlier than 2 months after the last potential exposure[†])

Serology[†]

+ Eosinophile count[‡]

(± urine POC-CCA testing[§] [always + routine urine dipstick testing for WBC and RBC])

In the case of a positive serology result, perform stool and urine microscopy as in symptomatic persons (see below).

II. Testing of symptomatic persons

Microscopy* of 3 urine^Δ and 3 stool samples for eggs (depending on the geographic prevalence of the different *Schistosoma* spp.; maps see p. 185)

+ Serology[†]

+ Eosinophile count[‡]

(+ urine POC-CCA testing[§] [always + routine urine dipstick testing for WBC and RBC])
(± blood PCR)[¶]

[†] As the sensitivity of serology is superior to microscopy and the presence of eosinophilia, serology is the primary screening tool [1]. **Serological assessment should include at least 2 different *schistosoma* specific assays.** Serological screening **should not be done earlier than 2 months after the last contact to potentially infectious fresh water**, to allow for full development of the parasite and formation of antibodies to the adult stage (IgM levels peak at 12–16 weeks, IgG levels around 20 weeks after infection). In the case of positive serology also consider cross-reactivity; see p.136.

[‡] Note that eosinophilia is not always present (travellers: ~50%, migrants: 15–30%). Especially in chronic infection (e.g. endemic populations and migrants) eosinophilia is frequently absent or may be attributable to other causes (e.g. strongyloides or filarial infection etc.).

[§] Antigen detection is indicative of active infection/treatment failure and would thus be more useful than serology which may remain positive for years even after successful treatment. The diagnostic role and sensitivity of circulating antigens (CCA, CAA) still remains to be defined, especially in asymptomatic patients with a low parasite burden (e.g. travellers). Currently, only one point-of-care (POC)/rapid-diagnostic-test for schistosomal antigen detection is commercially available: a urine CCA (Circulating Cathodic Antigen) cassette test [Rapid Medical Diagnostics, South Africa; <http://www.rapid-diagnostics.com>] with a sensitivity of 70–100% (largely depending on the intensity of infection) and a specificity of ~95%, according to the manufacturer). **Comment:** The highest concentrations of CCA are detected in *S. mansoni* infections and, therefore, the test is particularly useful to diagnose *S. mansoni* infections. CCA levels in *S. haematobium* infections are variable and appear to differ between regions, which lowers sensitivity. **False-positive results due to urinary tract infection, haematuria, and pregnancy have been reported.** This is primarily attributed to the fact that the polysaccharide structure of CCA contains repeating units of Lewis-X trisaccharide, a molecule which is a common epitope on human cells (especially anti-inflammatory cells like granulocytes). **Therefore, always perform parallel testing of the urine with a routine dipstick test to exclude asymptomatic urinary tract infection and/or haematuria!** A recent study showed that the combination of serology + urine POC-CCA testing is the most sensitive screening option for asymptomatic *S.mansoni* infection in Eritrean refugees and superior to stool microscopy [4]. In travellers, the role of the commercially available urine POC-CCA test for screening is currently unclear. Our own experiences suggest that specificity is problematic in this population and that positive results should be interpreted with caution.

* In infected travellers, microscopy has a sensitivity of only ~60%, as the parasite load is mostly very low. Therefore, investigate a minimum of 3 stool and 3 urine samples (collected on different days) and additionally use serology (± urine POC-CCA ± PCR).

^Δ **The recommended time for collecting urine is from 10:00 a.m. to 2:00 p.m. [2,3]. Collect as much urine as possible.** Whether the traditionally recommended physical exercise before collecting urine samples ('exercise urine' [e.g. hopping in place, running up and down the stairs before voiding urine]) increases the sensitivity of urine microscopy has recently been questioned [5].

[¶] PCR is an emerging diagnostic tool, especially in acute schistosomiasis [6]. Sensitivity and specificity may be superior to conventional diagnostic methods. However, if used for follow-up, it must be considered that parasite-DNA may circulate for several months after successful treatment.

- [1] Bierman WF et al. Presentation and diagnosis of imported schistosomiasis: relevance of eosinophilia, microscopy for ova, and serology. *J Travel Med* 2005;12:9-13.
- [2] Doehring E et al. Day-to-day variation and circadian rhythm of egg excretion in urinary schistosomiasis in the Sudan. *Ann Trop Med Parasitol* 1983;77:587-94.
- [3] Doehring E et al. Circadian variation of ova excretion, proteinuria, hematuria, and leukocyturia in urinary schistosomiasis. *Kidney Int* 1985;27:667-71.
- [4] Chernet A et al. Accuracy of diagnostic tests for *S. mansoni* infection in asymptomatic Eritrean refugees: serology and POC-CCA against stool microscopy. *CID* 2017;65:568-74.
- [5] Coulibaly JT et al. *Schistosoma haematobium* egg excretion does not increase after exercise: implications for diagnostic testing. *Am J Trop Med Hyg* 2018;98:772-5.
- [6] Soentjens P et al. Diagnosing acute schistosomiasis. *Clin Infect Dis* 2014;58:304-5.

Work-up of schistosomiasis patients suspected to be infected >1 year:

1. Assess liver (GOT, GPT, AP, albumin, INR) and kidney (creatinine or cystatin C) function
2. Check for concomitant chronic viral hepatitis B/C infection in patients from endemic regions
3. Sonographical assessment (hepatoportal pathology? [see p. 105]; genito-urinary pathology? [if present rule out bladder malignancy and urogenital tuberculosis]).
4. Women: gynaecological assessment to rule out concomitant genital involvement (vulvovaginal ulcer, fibrotic nodules ['sandy patches'])

Follow-up after treatment of schistosomiasis:

I. Parasitologically confirmed schistosomiasis (= eggs found at initial assessment):

1. Check 3 urine and/or 3 stool samples for eggs at 3, 6 and 12 months after treatment*
2. If eosinophilia was initially present, check differential blood count at 3 and 6 months after treatment†
3. (If urine POC-CCA was initially positive, consider repeating urine POC-CCA testing after 3 months§)
4. If initially present, follow-up hepatic and/or urogenital pathologies by imaging

II. Serologically diagnosed schistosomiasis:

1. If no eosinophilia was initially present, consider checking the eosinophil count at 7 days† (and if then positive, at 3 and 6 months) after treatment
 2. If eosinophilia is present, check differential blood count at 3 months and 6 months after treatment; eosinophilia should be cleared at 6 months after treatment
 3. If initially present, follow-up hepatic and/or urogenital pathologies by imaging
- * Because praziquantel is not ovicidal and eggs must migrate through host tissue to reach the stool/urine, egg shedding can persist for some time even after successful treatment. Eggs become avial within 3 months after being laid. Vitality of eggs can be assessed by microscopy (‡ a hatching test). Detection of vital eggs \geq 3 months after treatment indicates treatment failure or reinfection.

§ Specificity of urine POC-CCA is considerably lower than its sensitivity. Thus, we recommend not generally using urine POC-CCA for follow-up.

† Eosinophil counts may transiently rise just after praziquantel therapy (which would support the suspected diagnosis in only serologically diagnosed cases). In these cases, normalization of eosinophil count is sometimes slow. In case of a persistently elevated eosinophil count consider an alternative aetiology and screen for strongyloidiasis (\pm other parasites).

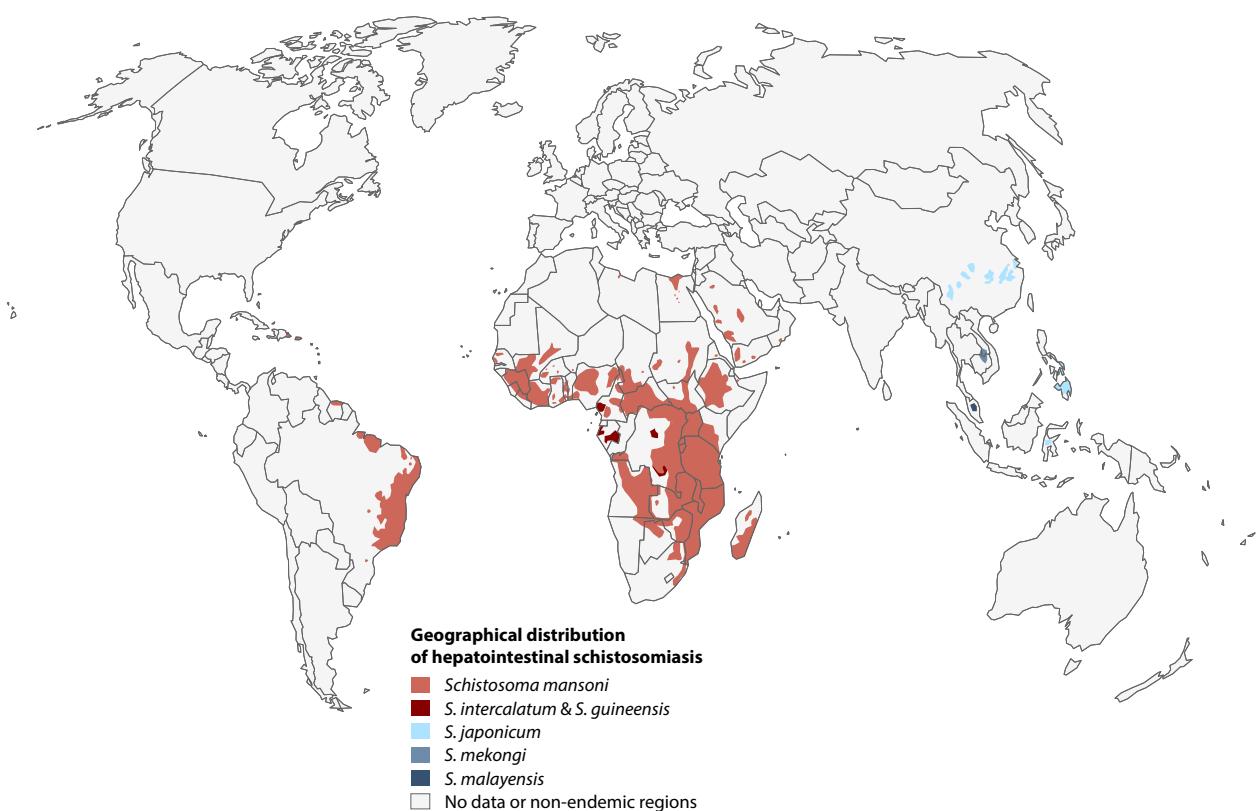
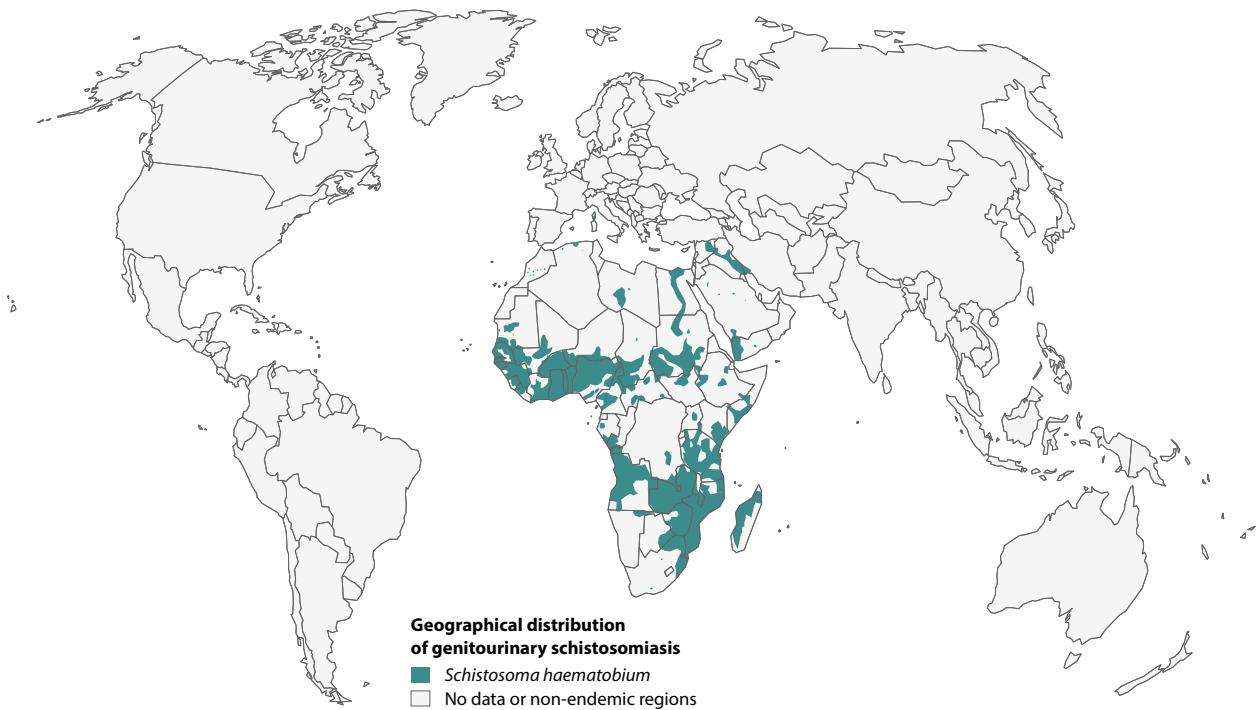
‡ Serological assays remain positive for prolonged periods (frequently up to several years) and may show a fluctuating titer pattern over time. Therefore, unlike other helminth serologies (e.g. strongyloides, filaria, etc.), serological follow-up has no major role in post-treatment monitoring of schistosomiasis.

Diagnostic pit-falls in schistosomiasis:

- False-positive *P. falciparum* histidine-rich protein 2 (HRP-2) immune-capture assay results may be caused by acute schistosomiasis due to *S. mekongi* [Lessem E et al. False-positive *P. falciparum* histidine-rich protein 2 immunocapture assay results for acute schistosomiasis caused by *S. mekongi*. *J Clin Microbiol* 2011;49:2331-2].
- Serology: be careful in the case of the following constellation:
 - Schistosoma egg-antigen ELISA: very high positive
 - Schistosoma adult-antigen ELISA: negative – low positive
 - IFAT: negative

Epidemiology of schistosomiasis: Map see p. 185.

Schistosomiasis



Human pathogenic *Schistosoma* spp.: *S. haematobium*: Africa, the Middle East, Corsica; *S. mansoni*: Africa, the Middle East, the Caribbean, Brazil, Venezuela, Suriname; *S. japonicum*: China, Indonesia, the Philippines; *S. mekongi*: Mekong river in Southern Laos (Si Phan Don region) and some districts in Cambodia [Muth S et al. *S. mekongi* in Cambodia and Lao PDR. *Adv Parasitol.* 2010;72:179-203]; *S. intercalatum* and related *S. guineensis*: Rain forest areas of central Africa; *S. malayensis*: closely related to *S. mekongi*, few transmission sites on the Malaysian Peninsula, infections confined to rural aboriginal communities [Latif B et al. Autochthonous human schistosomiasis, Malaysia. *EID* 2013;19:1340-11]; *S. mattheei* (not shown on map): primarily a zoonosis of cattle, human cases reported from South Africa. Schistosomiasis cases reported in Myanmar in 2018, species unknown [<http://www.promedmail.org/direct.php?id=20180329.5717098>].

Detailed maps providing epidemiological data on country level: http://www.who.int/schistosomiasis/epidemiology/global_atlas_maps/en/

Maps by Rosalie Zimmermann & Andreas Neumayr, adapted from WHO maps 92720 and 92721, Gryseels B. Lancet 2006;368:1106-1118 and IAMAT World Schistosomiasis Risk Chart 2015